

REVIEW ARTICLE – DEBATE ARTICLES: WHICH PET FLOW TRACER IS THE BEST FOR MBF QUANTIFICATION?

A New Era of Myocardial Perfusion Imaging: ^{18}F PET Tracer

Mitsuru Hirano, PhD¹⁾, Rudolf A. Werner, MD^{2), 3)} and Takahiro Higuchi, MD, PhD^{1), 2), 3)}

Received: March 1, 2018/Revised manuscript received: March 28, 2018/Accepted: April 4, 2018

J-STAGE advance published: July 31, 2018

© The Japanese Society of Nuclear Cardiology 2019

Abstract

Myocardial perfusion imaging (MPI) using advanced PET technology is increasingly used for non-invasive detection and evaluation of coronary artery disease (CAD), but is still limited for clinical use. Recently, ^{18}F labeled PET perfusion tracers have been actively developed as a novel class of PET MPI agents to overcome the disadvantages of conventional PET MPI tracers (^{15}O -labelled water, ^{13}N -ammonia, and ^{82}Rb chloride). This review summarizes the advantages and the feasibility of recent developed ^{18}F labeled tracers in clinical practice.

Keywords: ^{18}F -Flurpiridaz, ^{18}F labeled tracer, Coronary artery disease (CAD), Myocardial perfusion imaging (MPI), PET

Ann Nucl Cardiol 2019; 5 (1): 73–76

Currently, single photon emission computed tomography (SPECT) myocardial perfusion imaging (MPI) remains the mainstay of risk stratification of coronary artery disease (CAD). The most frequently employed radiotracers for SPECT MPI are ^{201}Tl , $^{99\text{m}}\text{Tc}$ -sestamibi and $^{99\text{m}}\text{Tc}$ -tetrofosmin. PET MPI is becoming an increasingly attractive alternative for clinical routine application. In principle, PET has a higher count sensitivity and higher temporal and spatial resolution than SPECT (1). ^{15}O -labelled water, ^{13}N -ammonia, and ^{82}Rb chloride are available for clinical use, but the routine clinical use of PET MPI is still limited, particularly due to the short half-life of conventional PET MPI tracers. The drawbacks of short half-life radiotracers inherit the requirement of either a cost-intensive generator or an onsite cyclotron for production. Furthermore, due to short half-lives, these tracers are only suitable for use in conjunction with pharmacologic stress, but not with physical exercise stress. Recently, ^{18}F labeled PET perfusion tracers have been actively developed as a new class of PET MPI agent to overcome these disadvantages. The longer half-life of ^{18}F , compared with other radionuclides (110 min for ^{18}F , 76 sec for ^{82}Rb chloride, 2.1 min for ^{15}O -labelled water, 10 min for ^{13}N -ammonia), may open avenues to make the tracer available as a unit dose from regional cyclotron

centers and allow the use in conjunction with treadmill exercise testing. The short positron range of ^{18}F (1.03 mm for ^{18}F , 8.6 mm for ^{82}Rb chloride, 4.14 mm for ^{15}O -labelled water, 2.53 mm for ^{13}N -ammonia) would result in considerably high spatial resolution.

Delivery of ^{18}F labeled tracers from regional centers

The radiotracers ^{15}O -labelled water, ^{13}N -ammonia, and ^{82}Rb chloride are clinically established for PET MPI. Because of a cyclotron dependency of ^{15}O -labelled water and ^{13}N -ammonia, their clinical use for PET MPI is limited to a few centers with on-site cyclotron (Figure 1a). This limitation could be overcome by the increasing use of ^{82}Rb chloride as it can be eluted from a commercially available ^{82}Sr generator (Figure 1b). However, the generator is expensive and must be replaced every month, limiting the use of ^{82}Rb chloride to large centers that have the volume of referrals to justify the cost (2). On the other hand, ^{18}F tracers with longer radioactive half-lives can be distributed by a central cyclotron facility in a manner similar to ^{18}F -fluoro-2-deoxy-D-glucose (FDG), allowing for the ordering of only the needed dosages per day (Figure 1c) (3).

doi: 10.17996/anc.18-00056

1) Department of Bio Medical Imaging, National Cardiovascular and Cerebral Research Center, Suita, Osaka, Japan

2) Department of Nuclear Medicine, University Hospital Würzburg, Würzburg, Germany

3) Comprehensive Heart Failure Center, University Hospital Würzburg, Würzburg, Germany

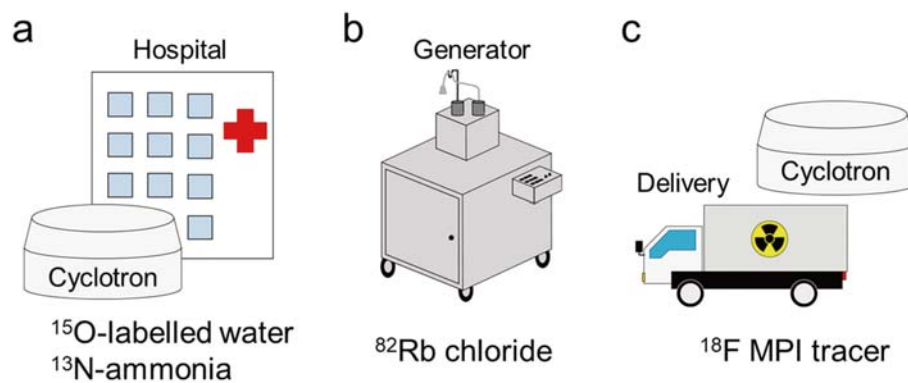


Figure 1 Schematic figure of required equipment for MPI tracers.

Physical exercise stress PET MPI

Rest and stress MPI is the most accurate test available for diagnosing CAD in patients who may be at risk for a heart attack at an early time of disease on-set. Rest and stress MPI have a proven risk-stratification capability and are sensitive to even the modest changes in coronary blood flow. Exercise is a safe physiologic stress, and exercise testing permits correlation of symptoms with exertion, measurement of exercise capacity and functional status, and assessment of efficacy of medical therapy (4). However, due to the short half-lives of current PET MPI tracers, stress imaging is only feasible in conjunction with pharmacologic stress. The half-life of ¹³N-ammonia (10 min) is longer than those of ¹⁵O-labelled water (2.1 min) and ⁸²Rb chloride (76 sec) and can be used in conjunction with supine bicycle exercise or treadmill exercise, but this approach is not practical for routine clinical use (5). Owing to the long half-life of ¹⁸F, ¹⁸F PET MPI tracer would be compatible with imaging protocols using exercise as well as pharmacologic stress. Actually, ¹⁸F-Flurpiridaz (¹⁸F-BMS-747158-02), which is the most promising ¹⁸F labeled tracer for MPI (6, 7), was used in conjunction with treadmill exercise in the previous clinical trials, and the feasibility of ¹⁸F-Flurpiridaz in rest and treadmill exercise has been demonstrated (8). It is known that a very high target-to-background ratio is noted when ¹⁸F-Flurpiridaz is injected at peak treadmill exercise. From the relationship between rest-stress contamination and dosing, it was determined that for a same-day rest-exercise protocol, a minimum dose ratio of 3.0 was needed, with a 60 min waiting time between the 2 injections. For an optimum same-day rest-adenosine stress protocol, on the other hand, a dose ratio of 3 with a 30 min waiting time between the 2 injections was required (5). In a pig model, single-scan method which provides quantitative rest-adenosine stress blood flow in less than 15 min was also suggested. A two-injection, single-scan protocol was used in which an adenosine infusion was started 4 min after the first injection of ¹⁸F-Flurpiridaz and followed either 3 or 6 min later by a second radiotracer injection (9).

The highest spatial resolution

The short positron range of ¹⁸F (1.03 mm) would result in the highest spatial resolution (3,5). ¹⁵O-labelled water has a 4.14 mm positron range resulting in an intermediate image resolution. The positron range of ¹³N-ammonia is 2.53 mm resulting in an intermediate-high image resolution. Due to the high energy of positrons emitted during the decay of ⁸²Rb chloride (positron range, 8.6 mm) and lower count rates as a result of the ultra-short half-life (76 sec), the spatial resolution of ⁸²Rb chloride is lower than those of other PET MPI tracers (10). As shown in Figure 2, the induced ¹⁸F-Flurpiridaz uptake defect corresponds precisely to the defect in ¹³N-ammonia images. Images obtained using ¹⁸F-Flurpiridaz demonstrated improved contrast and higher resolution resulting in better delineation of induced lesions, despite a higher injected dose of ¹³N-ammonia (57 MBq) versus ¹⁸F-Flurpiridaz (37 MBq) (11).

¹⁸F-Flurpiridaz as one of the most promising ¹⁸F PET MPI tracers and currently investigated in a Phase 3 trial in the United States

¹⁸F labeled PET perfusion tracers, can be divided into two categories: analogues of mitochondrial complex-1 (MC-1) inhibitors and lipophilic cations. Current research progress in development of MPI tracers has been well reviewed by Mou and Zhang (2017) (12). Especially, ¹⁸F-Flurpiridaz (¹⁸F-BMS-747158-02), which is one of analogues of MC-1 inhibitors, is the most promising ¹⁸F labeled tracer for MPI and the first of two Phase 3 trials have been completed (6, 7). ¹⁸F-Flurpiridaz was rapidly taken up into neonatal rat cardiomyocytes, and the half-time of washout was greater than 120 min, which can be considered as promising properties for perfusion imaging (13). The first-pass extraction fraction of ¹⁸F-Flurpiridaz by the myocardium is very high (>90% in isolated rat hearts perfused with the Langendorff method) and blood flow-independent, an important characteristic for stress myocardial blood flow (MBF) measurements (14). Furthermore, higher myocardial extraction facilitates detection of milder perfusion defects

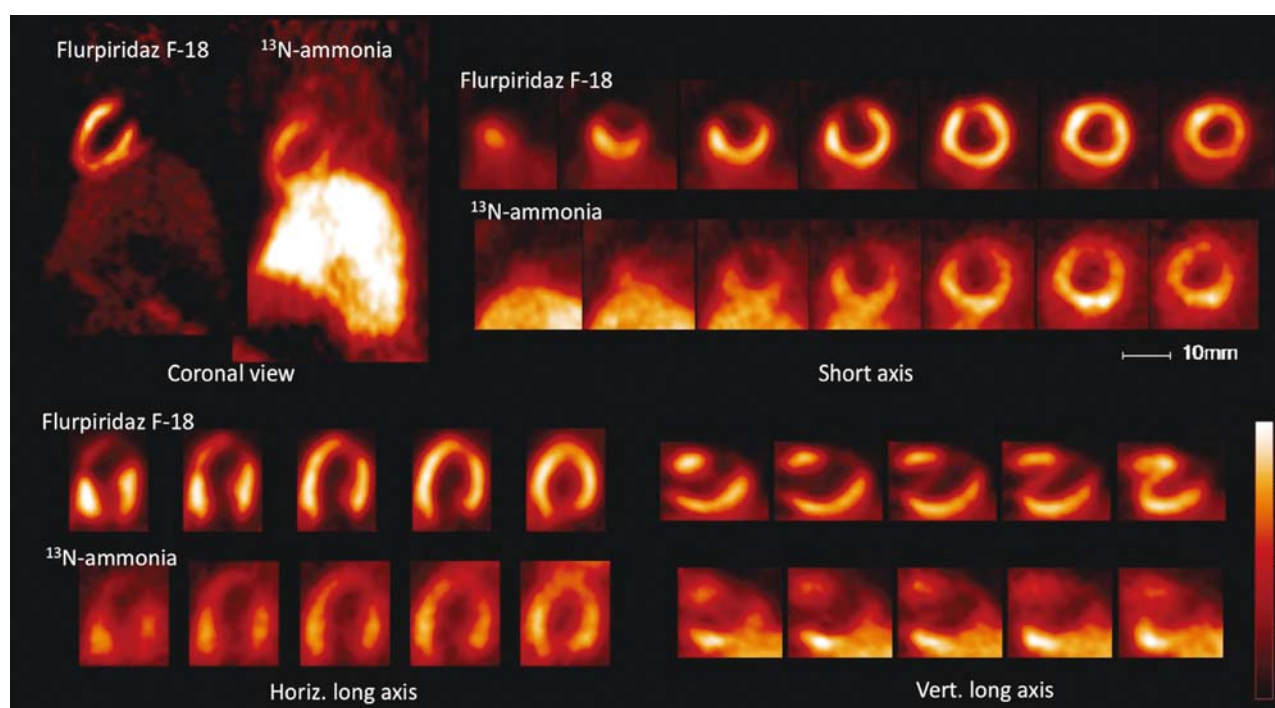


Figure 2 Coronal view, Short axis, Horizontal long axis, Vertical long axis images of a rat heart 1 week after coronary occlusion using ¹⁸F-Flurpiridaz PET and ¹³N-ammonia PET.

Table 1 Characteristics of various cardiac PET perfusion tracers

	¹⁸ F-Flurpiridaz	¹⁵ O-water	¹³ N-ammonia	⁸² Rb chloride
Half life	110 min	2.1 min	10 min	76 sec
Positron range	1.03 mm	4.14 mm	2.53 mm	8.6 mm
Production	Regional cyclotron	On-site cyclotron	On-site or nearby cyclotron	Generator
Image resolution	Highest	Intermediate	Intermediate-high	Lowest
Myocardial extraction fraction	>90%	100%	80%	65%
Treadmill exercise Imaging	Feasible	Not feasible	Feasible but not practical	Not feasible

(15). Absolute quantitation of MBF is important and is currently gaining ground in clinical practice (16). The elevated extraction fraction of ¹⁸F-Flurpiridaz at different flow rates makes it an optimal candidate for absolute MBF quantitation (5). In a rat biodistribution study, heart-to-lung and heart-to-liver ratios of ¹⁸F-Flurpiridaz were significantly higher than that of ^{99m}Tc-sestamibi SPECT (17). In a pig model, ¹⁸F-Flurpiridaz showed higher activity ratios between myocardium and blood, liver, and lungs compared with ¹³N-ammonia (18). In a Phase II clinical trial, ¹⁸F-Flurpiridaz had a higher diagnostic accuracy for evaluating multi-coronary artery stenosis, compared with SPECT MPI agents ^{99m}Tc-sestamibi, ^{99m}Tc-tetrofosmin, and ²⁰¹Tl (19). In a previous Phase 3 clinical trial, ¹⁸F-Flurpiridaz showed also a significant reduction in radiation exposure compared with SPECT. Of note, ¹⁸F-Flurpiridaz showed statistically superior sensitivity, specificity, accuracy, diagnostic confidence, and image quality compared with SPECT in obese subjects (6, 12).

Consequently, ¹⁸F-Flurpiridaz has entered Phase 3 multicenter clinical trials: A currently ongoing international study to evaluate the diagnostic efficacy of ¹⁸F-Flurpiridaz injection PET MPI in the detection of CAD will be completed in August 2020 (ClinicalTrials.gov Identifier: NCT03354273).

Future perspective

¹⁸F labeled PET MPI tracers offer several advantages due to their underlying physical properties (¹⁸F radioisotope) (Table 1). The use of PET MPI was mostly limited to large hospital due to the requirement for either onsite or very nearby cyclotrons as well as costly generators. However, similar to the successful commercialisation of ¹⁸F-FDG, ¹⁸F labeled PET MPI tracers can be produced in regional centers for dose-by-dose delivery to multiple PET sites. In addition to that, they may be also injected during exercise stress with enough time to move the patient to the camera after successfully performing the stress protocol (10). Moreover, ¹⁸F-Flurpiridaz

has also shown promising results in both preclinical examinations and clinical trials. Apart from that, other novel ¹⁸F labeled MPI tracers have been recently introduced (12). Hence, given their promising physical and imaging properties, ¹⁸F labeled MPI tracers might increase the acceptance of cardiac PET as a routine diagnostic tool.

Acknowledgments

None.

Sources of funding

None.

Conflicts of interest

None.

Abbreviation and Acronyms

CAD: coronary artery disease

MPI: myocardial perfusion imaging

FDG: ¹⁸F-fluoro-2-deoxy-D-glucose

MC-1: mitochondrial complex-1

MBF: myocardial blood flow

Reprint requests and correspondence:

Takahiro Higuchi, MD, PhD

Department of Bio Medical Imaging, National Cardiovascular and Cerebral Research Center, 5-7-1 Fujishiro-dai, Suita, Osaka 565-8565, Japan

E-mail: thiguchi@me.com

References

1. Higuchi T, Bengel FM. Cardiovascular nuclear imaging: from perfusion to molecular function: non-invasive imaging. *Heart* 2008; 94: 809–16.
2. Vitola JV, Delbeke D, eds. Nuclear cardiology and correlative imaging: a teaching file. Springer Science & Business Media, 2004.
3. Rischpler C, Park MJ, Fung GS, et al. Advances in PET myocardial perfusion imaging: F-18 labeled tracers. *Ann Nucl Med* 2012; 26: 1–6.
4. Chow BJ, Ananthasubramaniam K, de Kemp RA, et al. Comparison of treadmill exercise versus dipyridamole stress with myocardial perfusion imaging using rubidium-82 positron emission tomography. *J Am Coll Cardiol* 2005; 45: 1227–34.
5. Maddahi J, Packard RR. Cardiac PET perfusion tracers: current status and future directions. *Semin Nucl Med* 2014; 44: 333–43.
6. Bateman TM, Maddahi J, Udelson J, et al. Improved assessment of CAD in obese subjects with flurpiridaz F18 PET myocardial perfusion imaging: a subset analysis of the flurpiridaz F18 301 phase 3 study. *JACC* 2016; 67 suppl: 1578.
7. Heller GV, Maddahi J, Udelson JE, et al. Abstract 18022: Improved assessment of CAD in women with flurpiridaz F-18 PET myocardial perfusion imaging: results of subset analysis of the flurpiridaz F-18 phase 3 study. *Circulation* 2015; 132: A18022.
8. Maddahi J, Bengel F, Huang SC, et al. Phase 1 rest-stress study of F-18 labeled BMS747158 myocardial perfusion PET tracer: Human safety, dosimetry, biodistribution, and myocardial imaging characteristics. *J Nucl Med* 2009; 50 suppl 2: 184.
9. Guehl NJ, Normandin MD, Wooten DW, et al. Single-scan rest/stress imaging: validation in a porcine model with 18F-Flurpiridaz. *Eur J Nucl Med Mol Imaging* 2017; 44: 1538–46.
10. Bengel FM, Higuchi T, Javadi MS, et al. Cardiac positron emission tomography. *J Am Coll Cardiol* 2009; 54: 1–15.
11. Higuchi T, Nekolla SG, Huisman MM, et al. A new 18F-labeled myocardial PET tracer: myocardial uptake after permanent and transient coronary occlusion in rats. *J Nucl Med* 2008; 49: 1715–22.
12. Mou T, Zhang X. Research progress on 18F-labeled agents for imaging of myocardial perfusion with positron emission tomography. *Molecules* 2017; 22: 562.
13. Yalamanchili P, Wexler E, Hayes M, et al. Mechanism of uptake and retention of F-18 BMS-747158-02 in cardiomyocytes: a novel PET myocardial imaging agent. *J Nucl Cardiol* 2007; 14: 782–8.
14. Huisman MC, Higuchi T, Reder S, et al. Initial characterization of an 18F-labeled myocardial perfusion tracer. *J Nucl Med* 2008; 49: 630–6.
15. Maddahi J. Properties of an ideal PET perfusion tracer: new PET tracer cases and data. *J Nucl Cardiol* 2012; 19 Suppl 1: S30–7.
16. Gould KL, Johnson NP, Bateman TM, et al. Anatomic versus physiologic assessment of coronary artery disease. Role of coronary flow reserve, fractional flow reserve, and positron emission tomography imaging in revascularization decision-making. *J Am Coll Cardiol* 2013; 62: 1639–53.
17. Yu M, Guaraldi MT, Mistry M, et al. BMS-747158-02: a novel PET myocardial perfusion imaging agent. *J Nucl Cardiol* 2007; 14: 789–98.
18. Nekolla SG, Reder S, Saraste A, et al. Evaluation of the novel myocardial perfusion positron-emission tomography tracer 18F-BMS-747158-02: comparison to 13N-ammonia and validation with microspheres in a pig model. *Circulation* 2009; 119: 2333–42.
19. Berman DS, Maddahi J, Tamarappoo BK, et al. Phase II safety and clinical comparison with single-photon emission computed tomography myocardial perfusion imaging for detection of coronary artery disease: flurpiridaz F 18 positron emission tomography. *J Am Coll Cardiol* 2013; 61: 469–77.